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Editorial

Unlocking the role of liver sinusoidal endothelial cells: Key players in liver fibrosis: Editorial on “Liver sinusoidal endothelial cell: An important yet often overlooked player in the liver fibrosis”

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Liver sinusoidal endothelial cells (LSECs) are a critical component of the liver's microenvironment, constituting approximately 90% of the liver endothelial cell population.^{1,2} They play a vital role in liver homeostasis by regulating vascular tone, blood filtration, nutrient exchange, and immune modulation. However, in the context of liver fibrosis, which can progress to cirrhosis and cause serious complications such as portal hypertension, the function of LSECs becomes compromised. This dysfunction contributes significantly to the pathogenesis of liver fibrosis.

The review article by Qu et al.³ comprehensively discussed how LSECs contribute to liver fibrosis development and highlighted the molecular mechanisms underlying LSEC dysfunction. The authors also discussed how LSECs influence functions of hepatic stellate cells (HSCs), hepatocytes and immune cells, leading to inflammation and fibrosis. While previous research has focused heavily on HSCs

and hepatocytes as major players of fibrogenesis, the role of LSECs has been overlooked. LSEC dysfunction disrupts normal liver microcirculation, impairs liver regeneration, and promotes inflammation and fibrogenesis through dysregulated signaling pathways involving cytokines, chemokines, and angiogenic factors. Given the fact that there are no FDA approved anti-fibrotic drugs for liver fibrosis, understanding LSEC biology holds significant promise for a potentially novel therapeutic target for liver fibrosis.

LSEC dysfunction is associated with structural changes such as loss of fenestrae and capillarization (i.e., development of basement membrane of LSECs). The mechanism for the maintenance of fenestrate remains to be fully understood. The loss of fenestrae diminishes LSECs' antigen-presenting capability, consequently impairing immune response regulation. This review article by Qu et al.³ succinctly summarizes various studies elucidating potential factors regulating fenestrae, such as cytoskeletal proteins. The contraction and dilatation of fenestrae in LSECs are regulated by actin cytoskeleton.⁴ It has been demonstrated

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that cytochalasin D, an F-actin-depolymerizing agent, predominantly dispelled the formation of large stress fibers in dedifferentiated LSECs, leading to the reformation of fenestrates and consequently mitigating liver fibrosis.⁵ Thus, the strategies targeting LSECs' dynamic remodeling processes, such as cytoskeletal regulation and maintenance of fenestrae, show promise in mitigating fibrosis.⁵

The mechanism underlying LSEC capillarization remains incompletely understood. Capillarization is often characterized by an upregulation of CD34 expression in LSECs, which in normal state do not express. It is reported that vascular endothelial growth factor (VEGF), produced by hepatocytes and HSCs, maintains the phenotype of LSECs.⁶ However, in cirrhotic livers, VEGF secretion is increased,⁷ suggesting that LSEC capillarization may result from disruption in downstream VEGF signaling rather than lack of VEGF itself. In fact, it was shown that there was significant downregulation of both VEGF receptor Kdr (VEGFR2) and co-receptor Nrp1 in peri-central area (Zone 3 LSECs) of cirrhotic mouse liver,¹ potentially contributing to LSEC capillarization.

Decreased nitric oxide (NO) production by endothelial isoform of NO synthase (eNOS) is also considered a key marker of endothelial dysfunction.⁸ This dysfunction precedes noticeable liver injury and affects not only hepatocytes but also HSCs, contributing to fibrogenesis. Restoring eNOS-derived NO production has been shown to inhibit HSC activation and maintain them in a quiescent state, thereby inhibiting fibrogenesis.⁶

The review article³ also touches upon the reduced scavenging capacity of LSECs in cirrhotic livers. LSECs are one of the most powerful scavengers in the body, playing an important role in clearance of wastes and pathogens in blood that originated from the gut and the systemic circulation.⁹⁻¹¹ This activity is related to their expression of various endocytosis receptor genes, including scavenger receptors (Scarb1, Scarb2, Stab1 and Stab2) and mannose receptor (Mrc1), as well as genes of related activities such as Fc gamma-receptor IIb2 (Fcgr2b/CD32b). It was found that there was downregulation of all these genes in cirrhotic livers, suggesting decreased endocytic and clearance ca-

pacities of LSECs.¹ This may make cirrhotic patients more susceptible to infection and systemic inflammation. Interestingly, all these endocytosis-related genes were also most downregulated in pericentral (Zone 3) LSECs in cirrhotic mice. The decreased endocytic capacity of LSECs may be associated with their capillarization as well, because decreased CD32b was also used as an indicator of LSEC capillarization in some studies.¹²

The heterogeneity of LSECs, a crucial aspect not addressed in this review article by Qu et al.³, plays a pivotal role in liver pathology. LSECs exhibit zonal-specific alterations within the liver, contributing to the progression of liver fibrosis and cirrhosis. These cells form a distinct subset of endothelial cells (ECs) unique to the liver, and recent advancements, particularly in single-cell analysis techniques, have unveiled their complex molecular heterogeneity. This deeper understanding has shed light on the molecular mechanisms underlying liver fibrosis and portal hypertension, emphasizing the significance of LSEC dysfunction accompanied by increased adhesion molecule expression and secretion of cytokines and chemokines in liver pathophysiology. Notably, a recent study employing single-cell sequencing in human-derived liver nonparenchymal cells from normal and cirrhotic patients identified two disease-specific EC populations, characterized by CD34⁺PLVAP⁺VWA1⁺ and CD34⁺ PLVAP⁺ACKR1⁺.¹³ The authors named them "scar-associated ECs". In cirrhotic mouse livers, there was a significant upregulation of CD34, PLVAP and ACKR1 in LSECs of all zones in cirrhotic livers.¹ These findings suggest a potential avenue for targeted therapy by focusing on LSECs expressing these specific genes, offering a more disease-specific approach to managing cirrhosis.

In conclusion, while various therapeutic approaches have demonstrated efficacy in preclinical studies, the translation of these findings into effective clinical treatments for cirrhosis remains challenging. Targeting LSECs specifically, possibly through the adeno-associated virus system or nanoparticle-based delivery systems, holds potential as a novel therapeutic avenue. It's also crucial to differentiate between capillarization and angiogenesis when targeting LSECs effectively for anti-fibrotic strategies.¹⁴ To date, no

Abbreviations:

LSECs, liver sinusoidal endothelial cells; HSCs, hepatic stellate cells; VEGF, vascular endothelial growth factor; NO, nitric oxide; eNOS, endothelial isoform of NO synthase; ECs, endothelial cells

perfect markers have been identified that distinguish LSECs from other vascular ECs,^{7,15} leading to the absence of established LSEC-specific Cre mouse models.^{7,16} Well-recognized LSEC markers in healthy livers include the surface receptor CD32b,¹⁷ C-Type Lectin Domain Family 4 Member G (CLEC4G),¹⁸ Lyve1,¹⁹ and Stab2.²⁰ Electron microscopy remains the gold standard for identifying LSECs based on their fenestrae. Furthermore, examining specific molecular markers of dysregulated LSECs in acute or chronic experimental liver disease models presents some challenges. Future research aimed at comprehensively understanding the intricate biology of LSECs in both healthy and diseased states will play a pivotal role in the development of targeted therapies tailored to LSECs and in elucidating their intricate interplay within the liver microenvironment. Such endeavors are indispensable for the advancement of effective treatments for liver fibrosis and its associated complications.

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Conflicts of Interest

The authors have no conflicts to disclose.

REFERENCES

1. Su T, Yang Y, Lai S, Jeong J, Jung Y, McConnell M, et al. Single-cell transcriptomics reveals zone-specific alterations of liver sinusoidal endothelial cells in cirrhosis. *Cell Mol Gastroenterol Hepatol* 2021;11:1139-1161.
2. McConnell MJ, Kostallari E, Ibrahim SH, Iwakiri Y. The evolving role of liver sinusoidal endothelial cells in liver health and disease. *Hepatology* 2023;78:649-669.
3. Qu J, Wang L, Li Y, Li X. Liver sinusoidal endothelial cell: An important yet often overlooked player in the liver fibrosis. *Clin Mol Hepatol* 2024;30:303-325.
4. Braet F, Muller M, Vekemans K, Wisse E, Le Couteur DG. Antimycin A-induced defenestration in rat hepatic sinusoidal endothelial cells. *Hepatology* 2003;38:394-402.
5. Di Martino J, Mascalchi P, Legros P, Lacomme S, Gontier E, Bioulac-Sage P, et al. Actin depolymerization in dedifferentiated liver sinusoidal endothelial cells promotes fenestrae formation. *Hepatol Commun* 2018;3:213-219.
6. Xie G, Wang X, Wang L, Wang L, Atkinson RD, Kanel GC, et al. Role of differentiation of liver sinusoidal endothelial cells in progression and regression of hepatic fibrosis in rats. *Gastroenterology* 2012;142:918-927.e6.
7. Poisson J, Lemoinne S, Boulanger C, Durand F, Moreau R, Valla D, et al. Liver sinusoidal endothelial cells: Physiology and role in liver diseases. *J Hepatol* 2017;66:212-227.
8. Iwakiri Y, Groszmann RJ. The hyperdynamic circulation of chronic liver diseases: from the patient to the molecule. *Hepatology* 2006;43(2 Suppl 1):S121-131.
9. Ganesan LP, Mohanty S, Kim J, Clark KR, Robinson JM, Anderson CL. Rapid and efficient clearance of blood-borne virus by liver sinusoidal endothelium. *PLoS Pathog* 2011;7:e1002281.
10. Malovic I, Sørensen KK, Elvevold KH, Nedredal GI, Paulsen S, Erofeev AV, et al. The mannose receptor on murine liver sinusoidal endothelial cells is the main denatured collagen clearance receptor. *Hepatology* 2007;45:1454-1461.
11. Hansen B, Longati P, Elvevold K, Nedredal GI, Schledzewski K, Olsen R, et al. Stabilin-1 and stabilin-2 are both directed into the early endocytic pathway in hepatic sinusoidal endothelium via interactions with clathrin/AP-2, independent of ligand binding. *Exp Cell Res* 2005;303:160-173.
12. Ohmura T, Enomoto K, Satoh H, Sawada N, Mori M. Establishment of a novel monoclonal antibody, SE-1, which specifically reacts with rat hepatic sinusoidal endothelial cells. *J Histochem Cytochem* 1993;41:1253-1257.
13. Ramachandran P, Dobie R, Wilson-Kanamori JR, Dora EF, Henderson BEP, Luu NT, et al. Resolving the fibrotic niche of human liver cirrhosis at single-cell level. *Nature* 2019;575:512-518.
14. Xu M, Xu HH, Lin Y, Sun X, Wang LJ, Fang ZP, et al. LECT2, a Ligand for Tie1, plays a crucial role in liver fibrogenesis. *Cell* 2019;178:1478-1492.e20.
15. Strauss O, Phillips A, Ruggiero K, Bartlett A, Dunbar PR. Immunofluorescence identifies distinct subsets of endothelial cells in the human liver. *Sci Rep* 2017;7:44356.
16. Payne S, De Val S, Neal A. Endothelial-specific cre mouse models. *Arterioscler Thromb Vasc Biol* 2018;38:2550-2561.
17. Mousavi SA, Sporstøl M, Fladeby C, Kjekken R, Barois N, Berg T. Receptor-mediated endocytosis of immune complexes in rat liver sinusoidal endothelial cells is mediated by FcγRIIb2. *Hepatology* 2007;46:871-884.

18. Aizarani N, Saviano A, Sagar, Maily L, Durand S, Herman JS, et al. A human liver cell atlas reveals heterogeneity and epithelial progenitors. *Nature* 2019;572:199-204.
19. Mouta Carreira C, Nasser SM, di Tomaso E, Padera TP, Boucher Y, Tomarev SI, et al. LYVE-1 is not restricted to the lymph vessels: expression in normal liver blood sinusoids and down-regulation in human liver cancer and cirrhosis. *Cancer Res* 2001;61:8079-8084.
20. McCourt PA, Smedsrød BH, Melkko J, Johansson S. Characterization of a hyaluronan receptor on rat sinusoidal liver endothelial cells and its functional relationship to scavenger receptors. *Hepatology* 1999;30:1276-1286.